



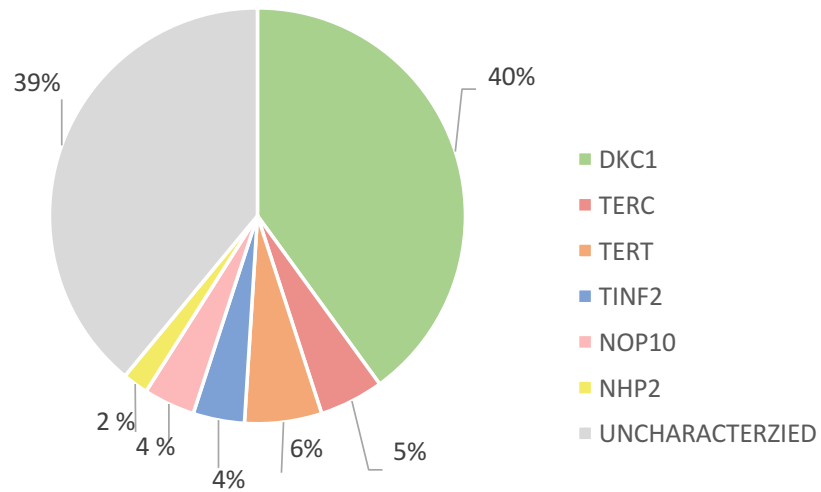
SAPIENZA
UNIVERSITÀ DI ROMA

Bone Marrow rescue in Dyskeratosis Congenita patients via autologous transplantation of gene-edited Hematopoietic (HSCs) and Mesenchimal Stem Cells (MSCs)

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Gene Therapy
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Background



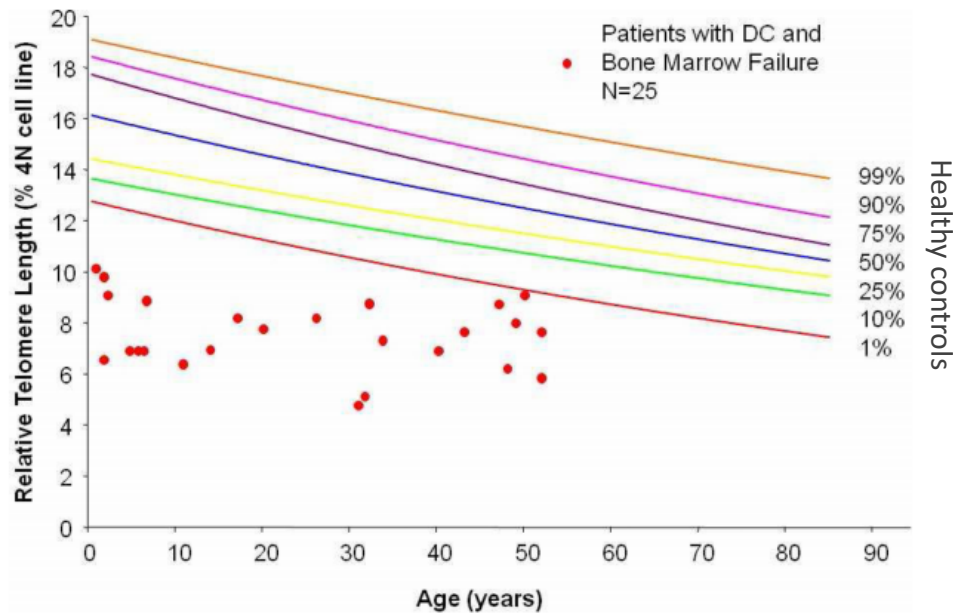
- Classical linkage analysis established the DKC1 gene as the gene responsible for the X-linked Dyskeratosis Congenita (DC).

(Dokal I. et al., Nature Genetics, 1998)

Family	Ethnic Origin	Exon	DNA Change	Protein Change
DCR-004	United Kingdom	11	1058C→T	A353V
DCR-008	United Kingdom	3	115A→G	K39E
DCR-006	Italy	11	1058C→T	A353V
DCR-055	Spain	11	1058C→T	A353V
DCR-009	Italy	10	961C→G	L321V
NIMGS 515	United States	4	196A→G	T66A
DCR-020	France	11	1058C→T	A353V
DCR-021	Austria	11	1050G→A	M350I
DCR-027	India	12	1204G→A	G402R
DCR-029	United States	11	1058C→T	A353V

(Knight S. W. Et al., Am. J. Hum. Genet.)

Background

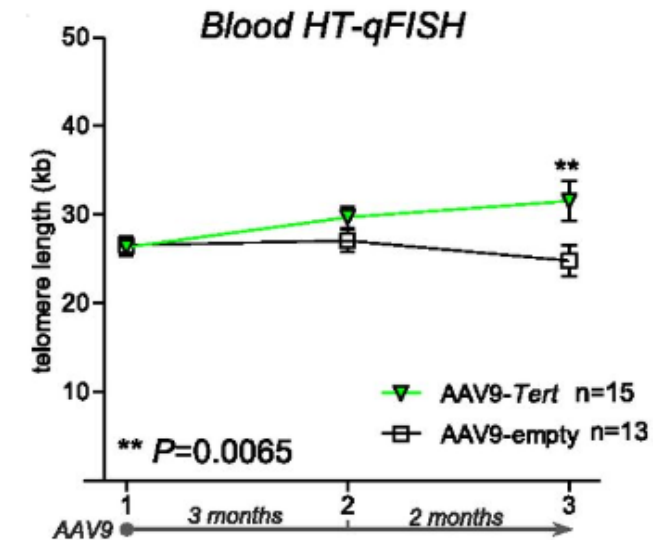


- **AAV9-Tert treatment** of AA mice rescued mortality due to Aplastic Anemia, concomitant with telomere re-elongation in blood and bone marrow cells.

(Bär C. et al., Blood, 2016)

- DC patients have very **short telomeres** in length compared with healthy age-matched controls

(Mason J. Et al., Cancer Genet., 2011)



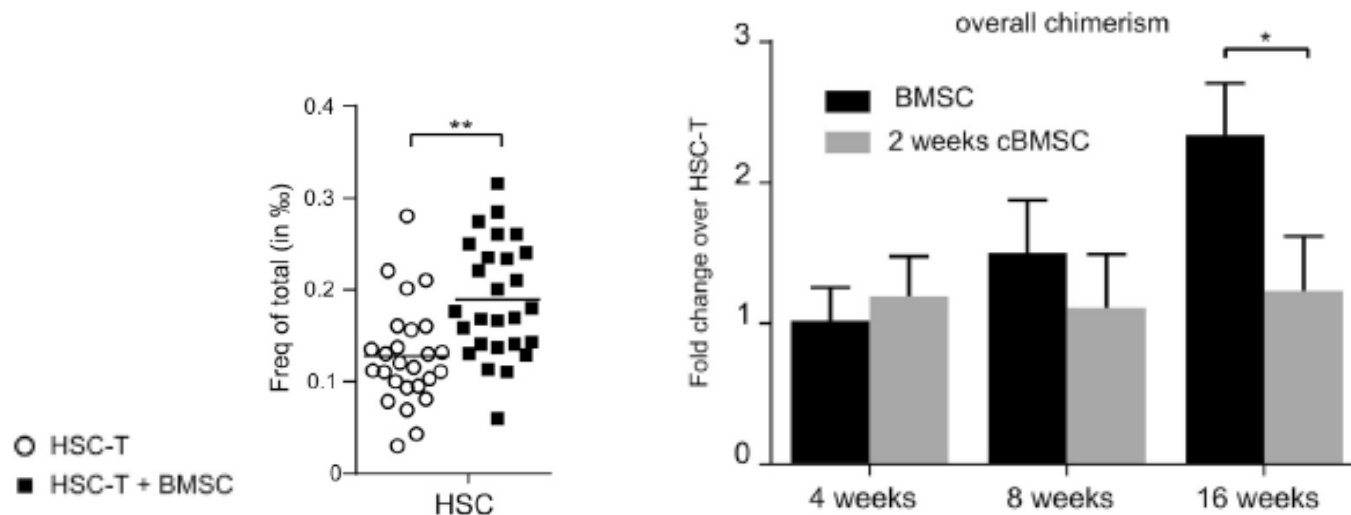
Background

- The only current interventions to treat these diseases involve **organ transplantation**; i.e., BM, liver, and lung

(Stanley S.E. et al., Curr Opin Genet Dev, 2015)

- Long-term (16 weeks) reconstitution assays confirmed expansion of functional HSCs in hosts **co-transplanted** with primary **BMSCs and HSCs** as compared to classical HSC-T alone.

(Abbuehl J.P. et al., Cell Stem Cell, 2017)



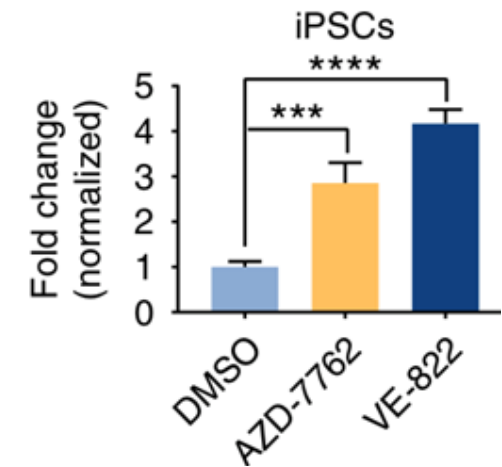
Background- toolkit

	Cas9	Cpf1
Components	crRNA, tracrRNA, Cas9	crRNA, Cpf1
Size	spCas9 4.2 kb	3.7 kb
gRNA length	100 nt	42 nt
tracrRNA	yes	no
dsDNA cleavage	Blunt end	5' overhang
PAM sequence	NGG	TTTN

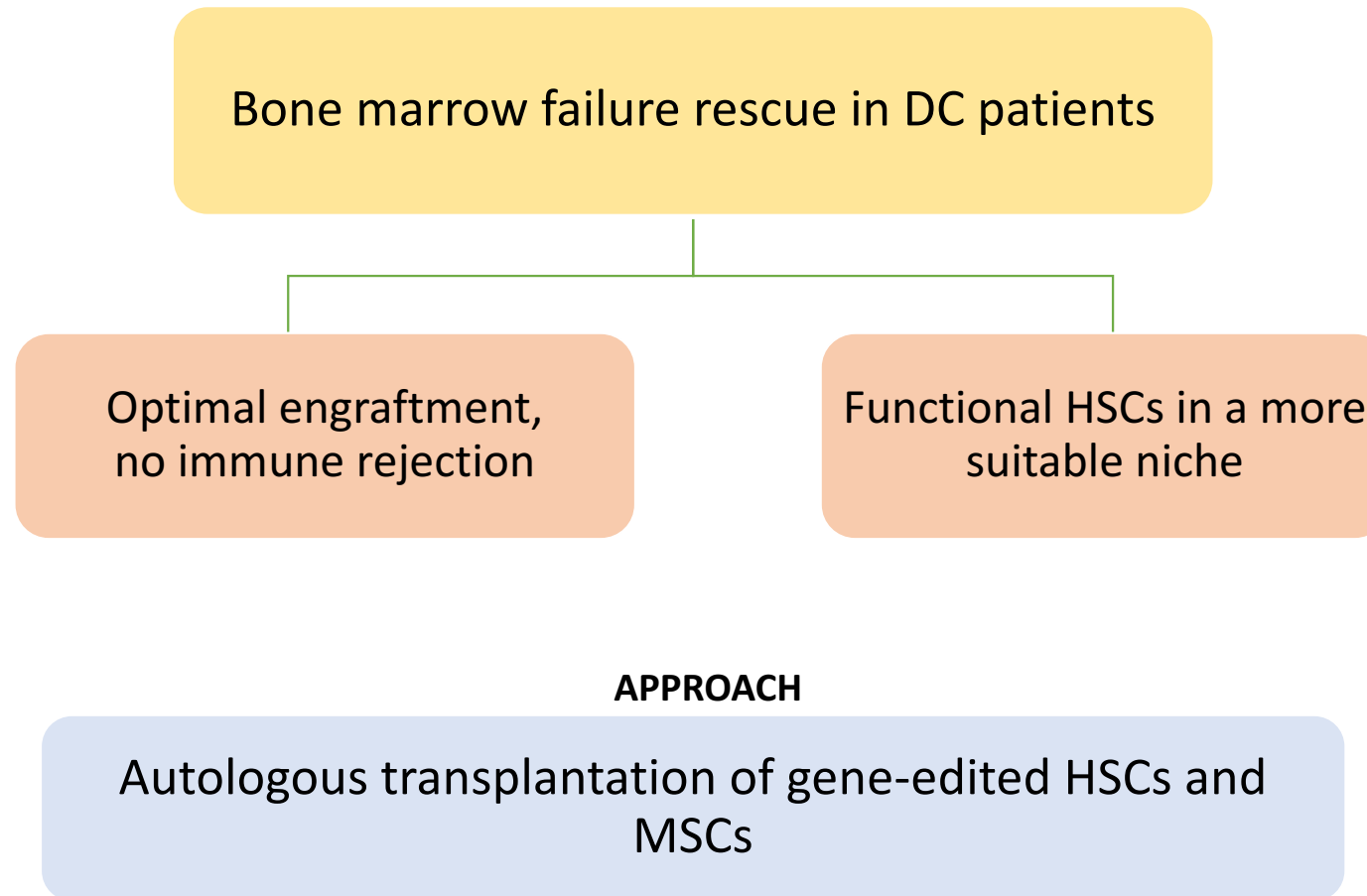
(Zetsche B. et al., Cell, 2015)

- **VE- 822** and **AZD-7762** significantly promoted CRISPR-Cpf1- mediated knock-in in hPSCs by 6-fold.

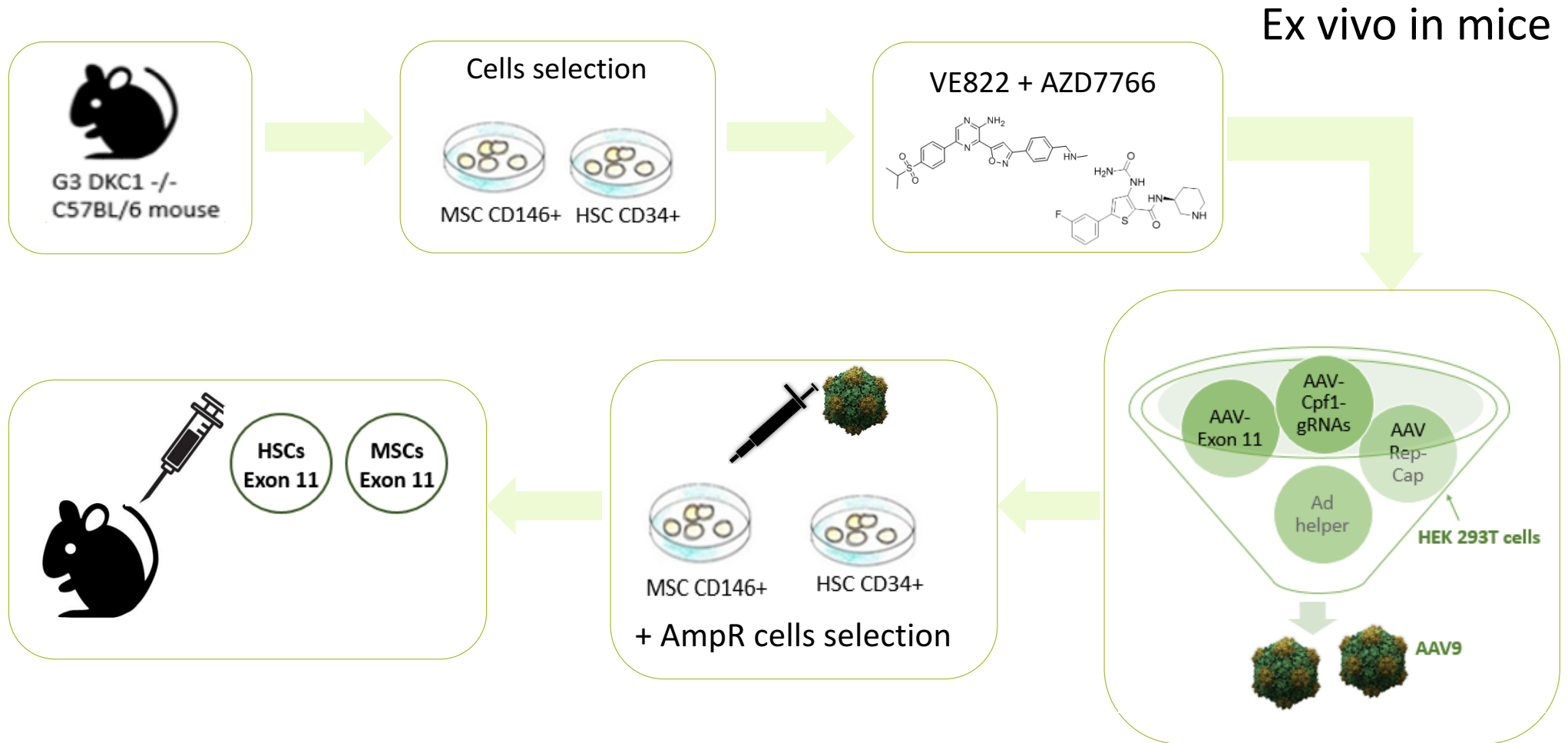
(Ma X. Et al., Nature Communications, 2018)



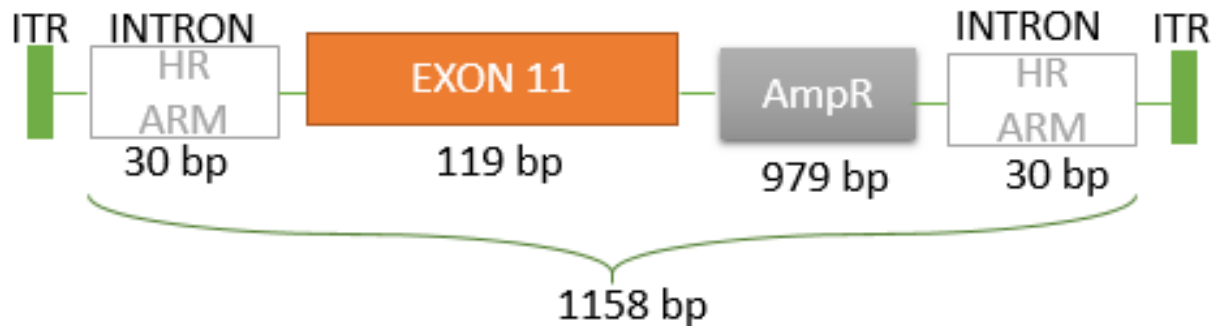
Objectives and strategy



Experimental plan – part one

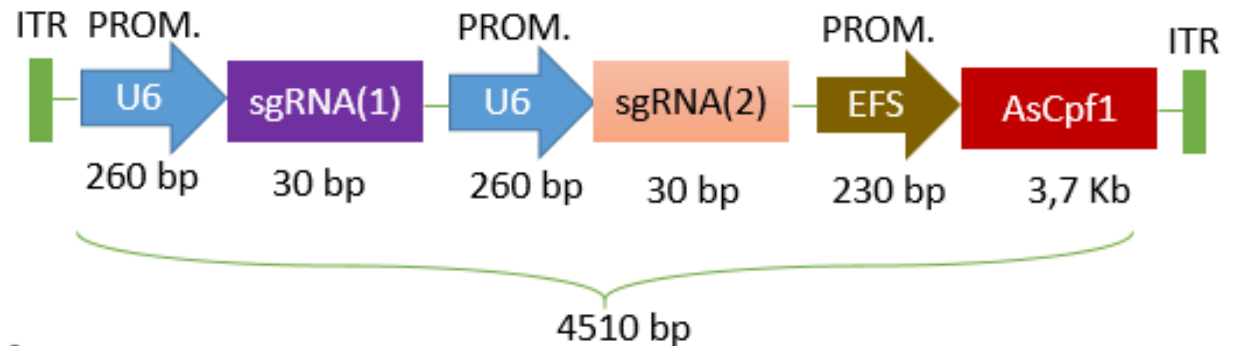


AAV vectors



AAV-9 vector

- non-integrative
- poor immunogenicity
- transduction in both dividing and quiescent cells
- long-term expression

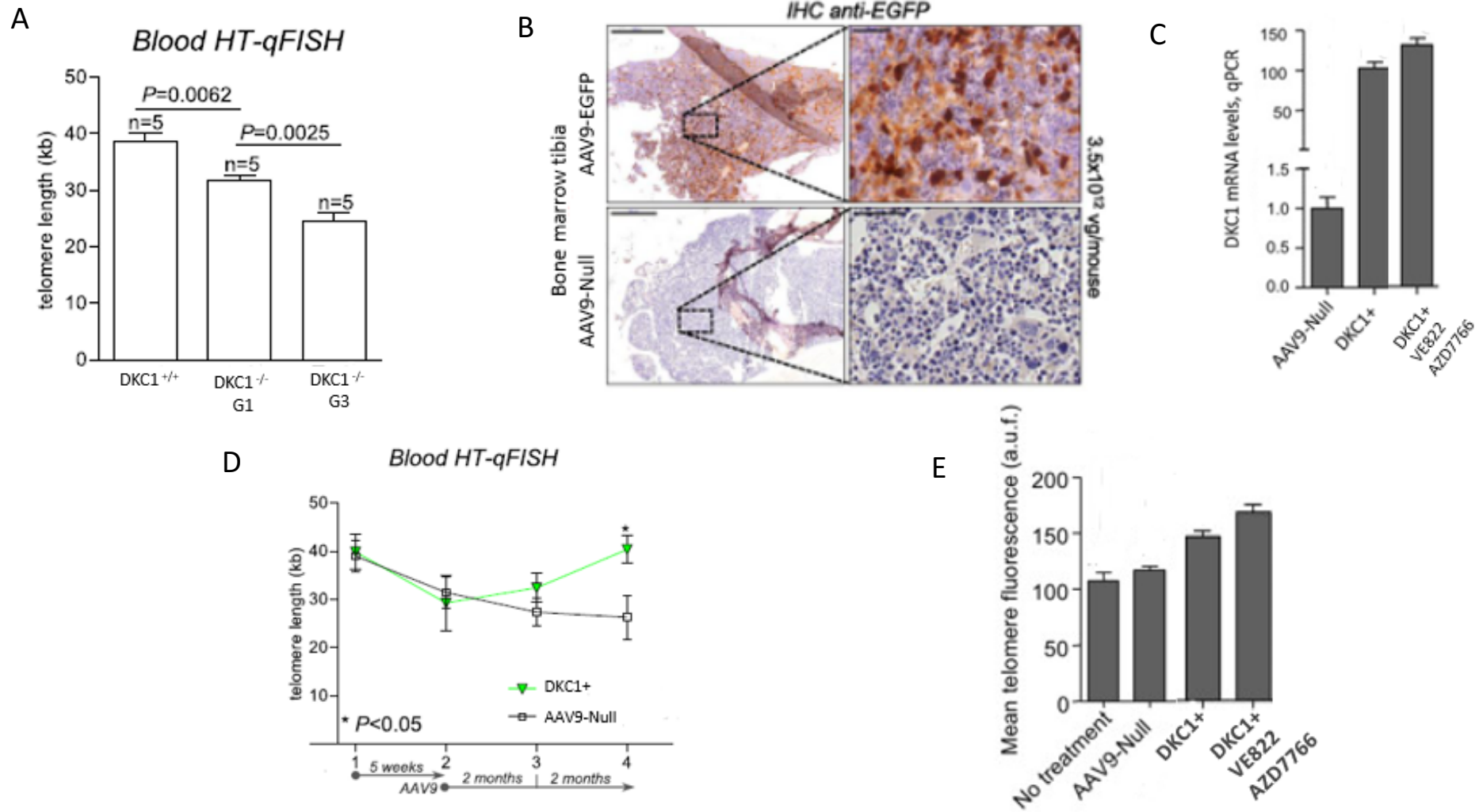


sgRNA(1) TTTTGAGTCGTGTGTAATGTGCGGAGAGTGGTAC

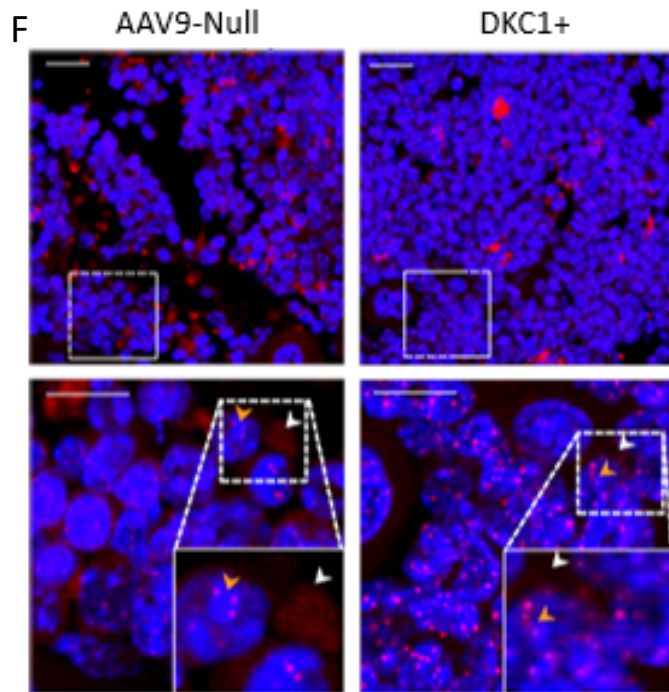
sgRNA(2) GCAGCTAGTGGGCTATAAGTGTCATCCCTGTTTC

(EuPaGDT.com)

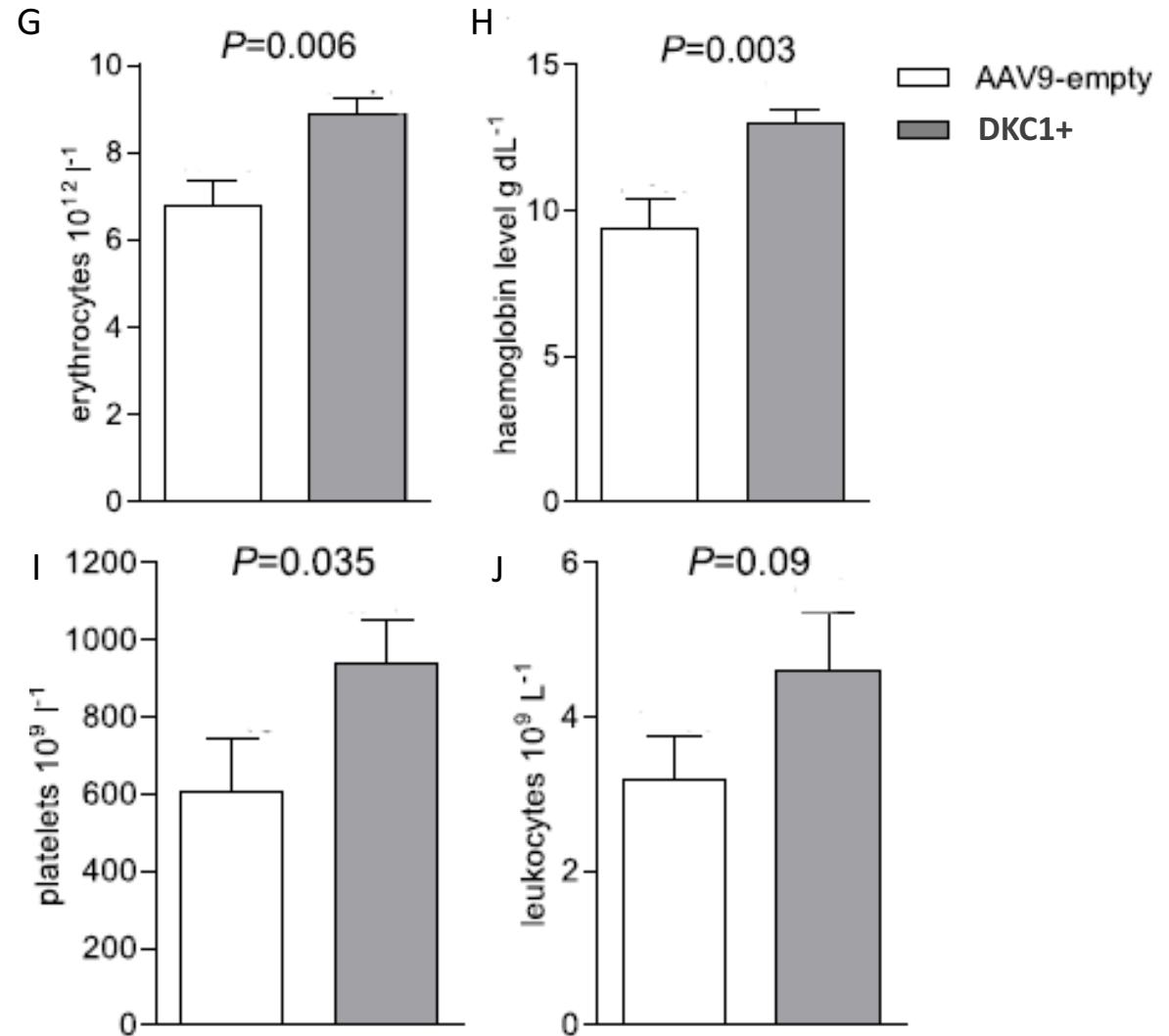
Expected results



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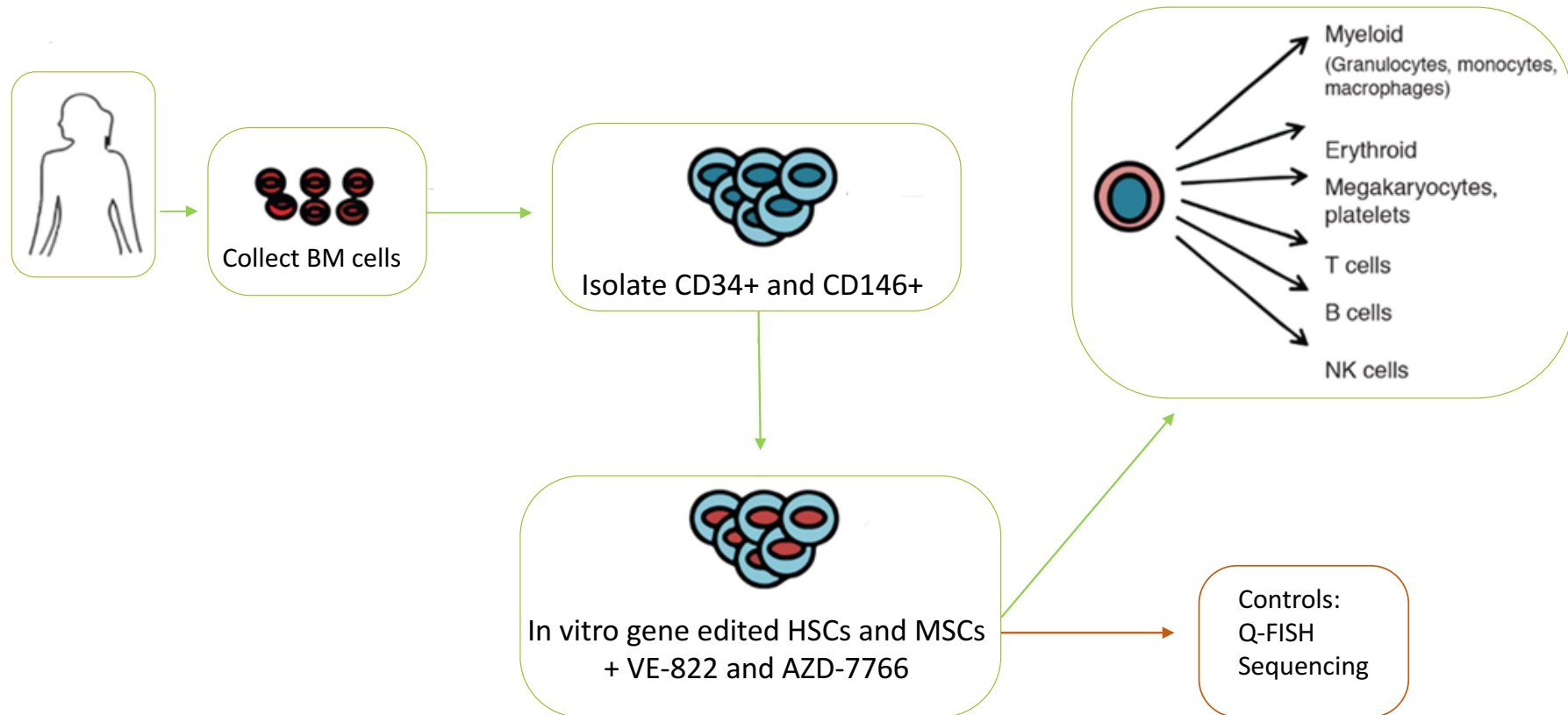


All control experiments to be performed at 4, 12 and 24 weeks



Experimental plan – part two

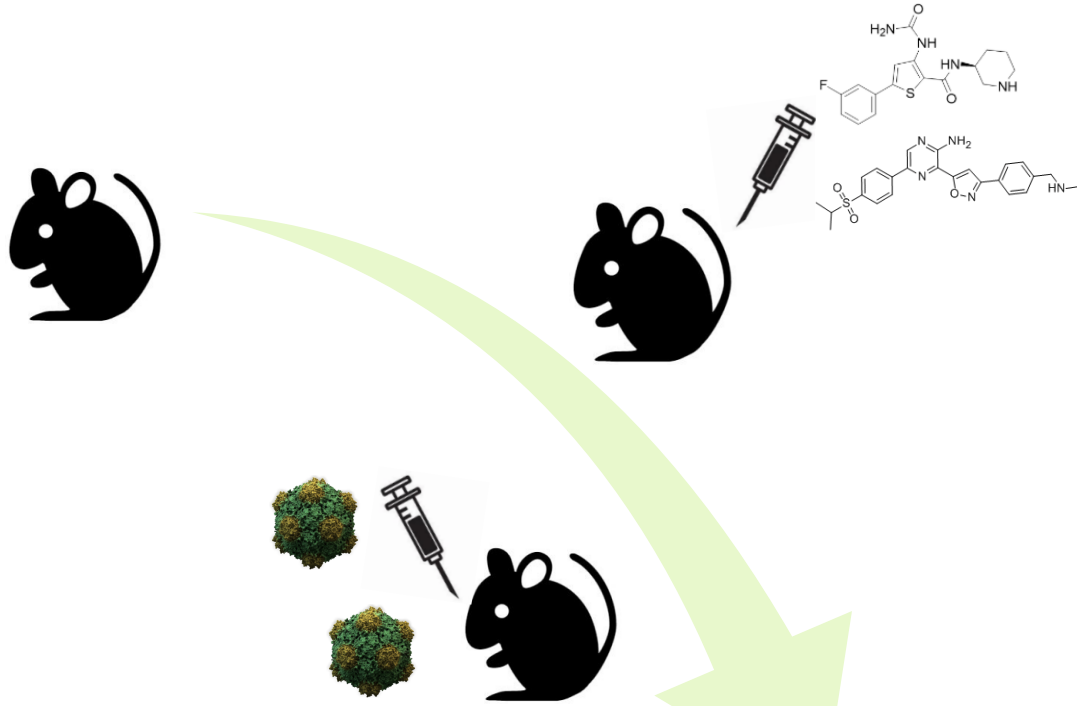
Ex vivo in humans



(Adapted from Jung M. et al., The American Society of Cell & Gene Therapy, 2015)

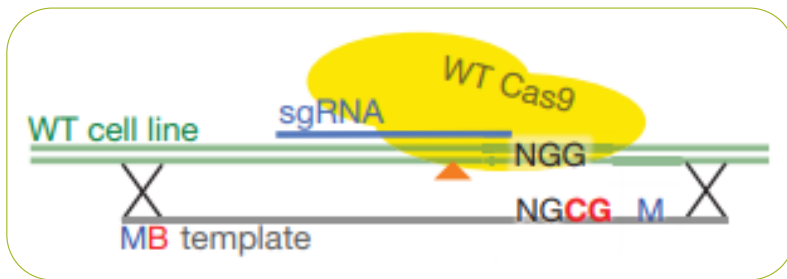
Experimental plan – part three

In vivo in mice



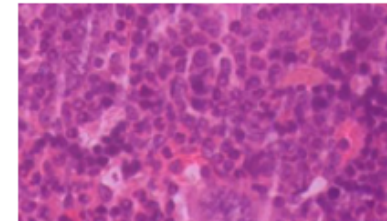
Crispr/Cpf1
attainment check

(Paquet et al., Nature, 2016)

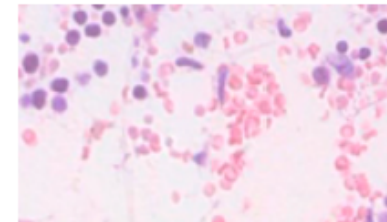


Expected results

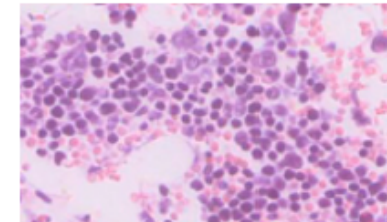
WT mouse



AAV9-Null



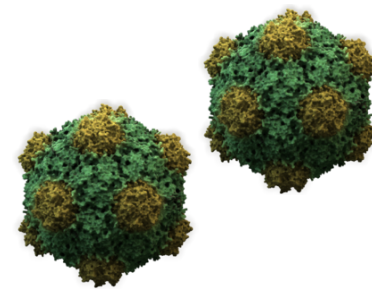
DKC1+



+ Sequencing

Ex-post Evaluation

PITFALLS	SOLUTIONS
Low packaging capacity	Lentiviral delivery
Lower VE-822 and AZD-7766 efficiency than expected	Dosage adjustment



MATERIALS	COSTS	SOURCE
C57BL/6J mouse models	20€ x 40 mice = 800€	thejacksonlaboratory.org
Stabulation	3000€	/
Antibodies	600€	ThermoFisher
VE822 (50 mg) + AZD7766 (50 mg)	240 + 360 € = 600€	MedChemExpress
r-AAV production		
HEK 293T cells	557€	Integrated DNA Technologies
Vector kit	500€	//
Crispr/Cpf1 + EFS promoter	480€	//
sgRNAs + U6 promoter	110€	//
FACS		
Amp selection kit	300€	Cellbiolabsinc.com
HT Q-FISH		
PBS	80€	Sigmaaldrich.com
Acetic acid	60€	//
Methanol	35€	//
DAPI	45€	//
96-well plates	900€	//
Telomere PNA FISH kits, PNA FISH kit, Cy3	1000€	Agilent.com
Immunohistochemistry kit	2000€	ThermoFisher
qPCR	300€	LifeTechnologies
Immunofluorescence kit	1500€	ThermoFisher
Blood cells count	300€	Analysis Laboratory
Sequencing	750€	Illumina

Required fundings: € 14.000
Experiment time span: 30 months

References

- Heiss N. S., Knight S. W., Vulliamy T. J., Klauck S. M., Wiemann S., Mason P. J., ... & Dokal I. (1998). X-linked dyskeratosis congenita is caused by mutations in a highly conserved gene with putative nucleolar functions. *Nature genetics*, 19(1), 32.
- Knight S. W., Heiss N. S., Vulliamy T. J., Greschner S., Stavrides G., Pai G. S., ... & Poustka A. (1999). X-linked dyskeratosis congenita is predominantly caused by missense mutations in the DKC1 gene. *The American Journal of Human Genetics*, 65(1), 50-58.
- Mason P. J., & Bessler M. (2011). The genetics of dyskeratosis congenita. *Cancer genetics*, 204(12), 635-645.
- Bär C., Povedano J. M., Serrano R., Benitez-Buelga C., Popkes M., Formentini I., ... & Blasco M. A. (2016). Telomerase gene therapy rescues telomere length, bone marrow aplasia and survival in mice with aplastic anemia. *Blood*, 127(14): 1770-1779.
- Stanley S. E., & Armanios M. (2015). The short and long telomere syndromes: paired paradigms for molecular medicine. *Current opinion in genetics & development*, 33, 1-9.
- Abbuehl J. P., Tatarova Z., Held W., & Huelsken J. (2017). Long-term engraftment of primary bone marrow stromal cells repairs niche damage and improves hematopoietic stem cell transplantation. *Cell stem cell*, 21(2), 241-255.
- Zetsche B., Gootenberg J. S., Abudayyeh O. O., Slaymaker I. M., Makarova K. S., Essletzbichler P., Volz S. E., Joung J., van der Oost J., Regev A., Koonin E. V., Zhang F. (2015). Cpf1 is a single RNA-guided endonuclease of a class 2 CRISPR-Cas system. *Cell*, 163(3), 759-71.
- Ma X., Chen X., Jin Y., Ge W., Wang W., Kong L., ... & Fu J. (2018). Small molecules promote CRISPR-Cpf1-mediated genome editing in human pluripotent stem cells. *Nature communications*, 9(1), 1303.
- Jung M., Dunbar C. E. and Winkler T. (2015). Modeling Human Bone Marrow Failure Syndromes Using Pluripotent Stem Cells and Genome Engineering, *The American Society of Gene & Cell Therapy*.
- Muñoz-Lorente M.A., Martínez P., Tejera Á., Whittemore K., Moisés-Silva A.C., Bosch F., Blasco M.A. (2018). AAV9-mediated telomerase activation does not accelerate tumorigenesis in the context of oncogenic K-Ras-induced lung cancer. *PLOS Genetics*, 14(8): e1007562.
- Paquet D., Kwart D., Chen A., Sproul A., Jacob S., Teo S., Olsen K.M., Gregg A., Noggle S. & Tessier-Lavigne M. (2016). Efficient introduction of specific homozygous and heterozygous mutations using CRISPR/Cas9. *Nature*, 533(7601):125-9.