

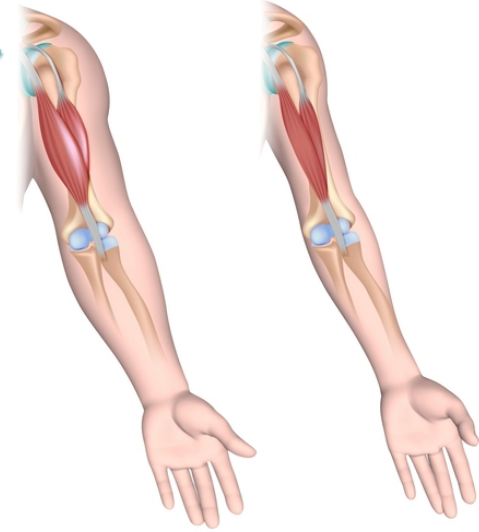
# Emery-Dreifuss muscular dystrophy: Silencing and restoring of LMNA gene

Gene therapy  
Professoressa I.Saggio  
2019/2020

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# Emery-Dreifuss muscular dystrophy

Normal biceps      Muscular dystrophy

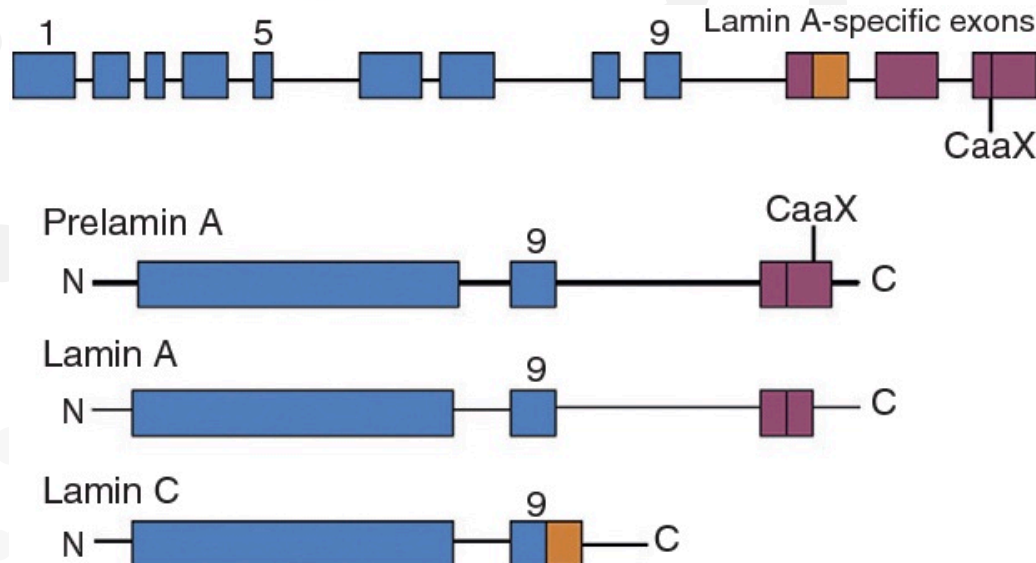


It is characterized by

- the triad of weakness of the shoulder and pelvic girdle muscles,
- contractures of the elbows, neck, and Achilles tendon,
- cardiac involvement, most commonly arrhythmias

## The mutation

EDMD2 is caused by heterozygous mutation in the gene encoding lamin A/C (LMNA on chromosome 1q22). 23 different mutations are distributed between exons 1 and 9 in the region of LMNA common to both lamins A and C.



LMNA  
gene

# Aims of the project

**1. SILENCING  
the ineffective LMNA  
gene**



**2. RESTORING  
the wild type  
phenotype**

**3. REDUCE  
off-targets events**

# Strategy and why



## Why third-generation lentivirus vector?

Advantages:

- High packaging capacity,
- Stable gene expression in both dividing and post-mitotic cells,
- Low immunogenicity in the recipient organism.

01

### SILENCING LMNA

CRISPR/Cas9 delivered using cell derived nanovesicles: GESICLES

## Why CRISPR /Cas9 gesicles?

- No persistent expression of Cas9;
- Glycoproteins on their surface mediate binding and fusion with skeleton muscles cells;
- Allows control of the dose and duration of the complex in the cell, reducing the chance of off-target effects.

### RESTORING WT LMNA

Third-generation lentivirus vector

02



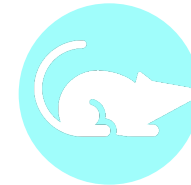
# Experimental plan

Years

1<sup>st</sup>

2<sup>nd</sup>

3<sup>rd</sup>



## Production of the vectors

Development of CRISPR/Cas9 gesicles and of third-generation lentivirus vectors with LMNA gene

## In vitro experiment part 1

CRISPR/Cas9 transfection with gesicles for silencing the endogenous LMNA gene expression

## In vitro experiment part 2

Third-generation lentiviral vectors delivery

## In vivo experiment

Transfection of neonatal mice mutant for LMNA gene (H222P mice)

## Results

Restoring of the WT phenotype and rescue of the muscles

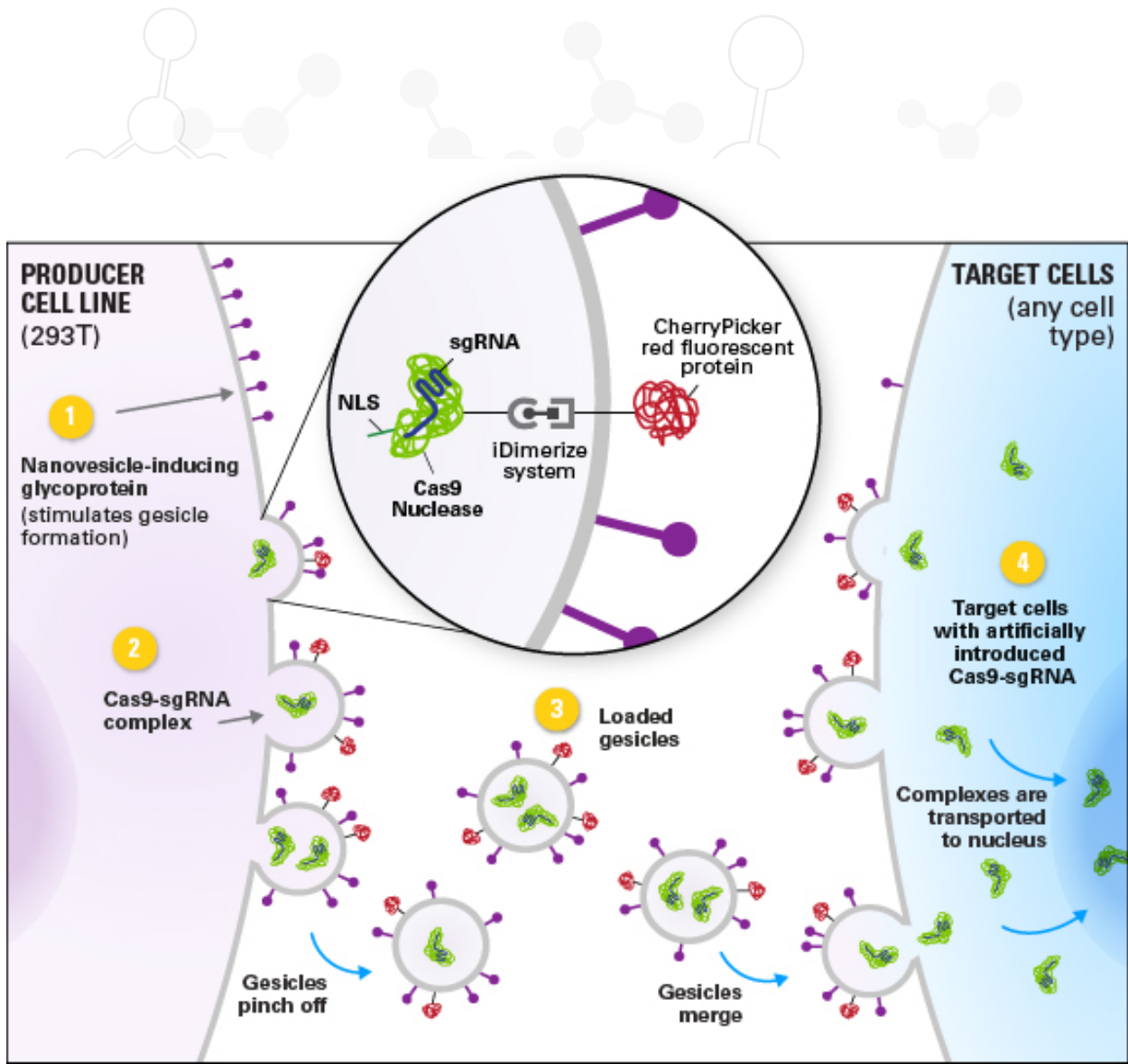


Figure B adapted from: "www.takarabio.com"

A

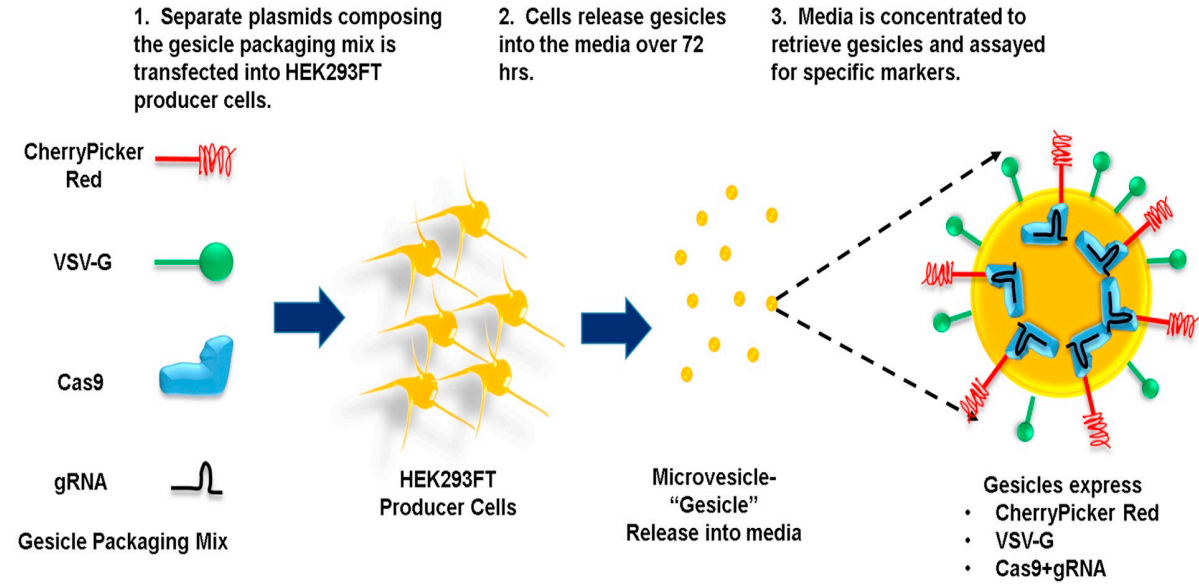


Figure A adapted from: "Lee A. Campbell, et al., 2019"

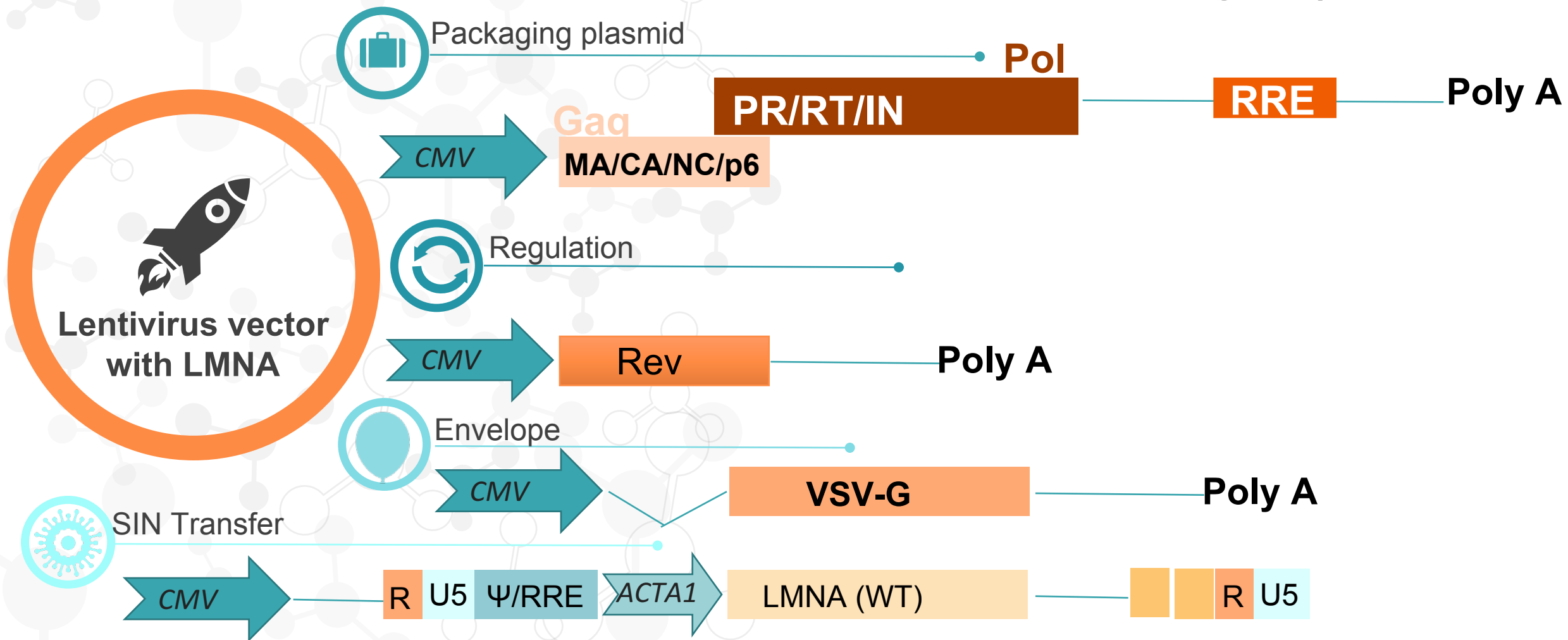
# Delivering CRISPR-CAS9 with vesicles

# Third-generation lentivirus vector

Using a specific promoter for skeleton muscles cells which is the one of Actin alpha 1 (ACTA1), which is expressed in skeletal muscle and target it with GFP fused to the gene. The generation of lentivirus vector will be in HEK cells.



1 .LV-MAX Lentiviral Production System protocol overview



# In vitro



**1<sup>st</sup> culture**

with sgRNA1

**2<sup>nd</sup> culture**

with sgRNA2

**3<sup>rd</sup> culture**

with sgRNA3



**4<sup>th</sup> culture**

Empty vector

**5<sup>th</sup> culture**

Cells from WT mice

## Three different **sgRNA**

Three different sgRNAs have been tested in order to establish the best one and use it for the in vivo experiments.

## Culture of skeleton-muscles cells from **H222P mice**

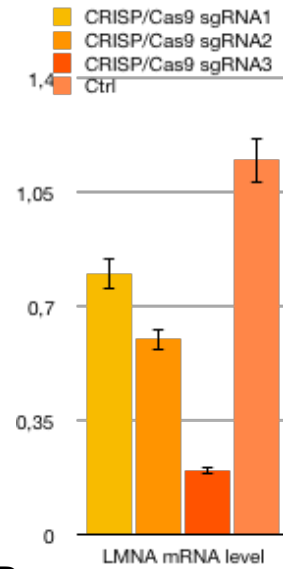
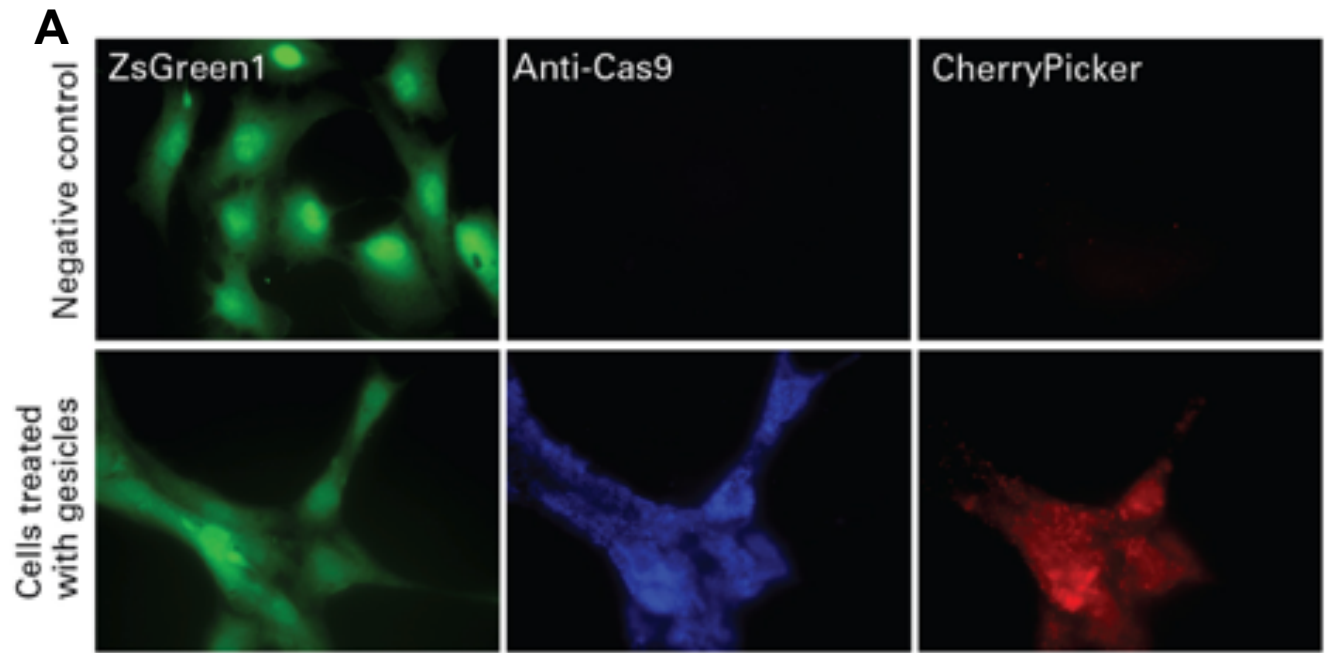
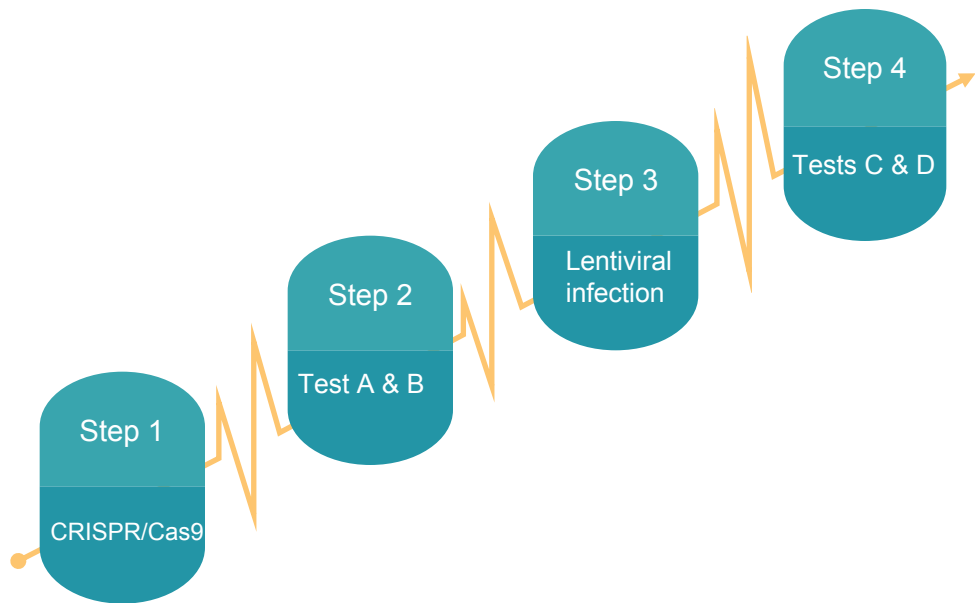
We chose the LMNA H222P missense mutation to create a faithful mouse model of this autosomal dominant Emery-Dreifuss muscular dystrophy.

## Culture with H222P cells treated with **sgRNA3** are injected with lentivirus

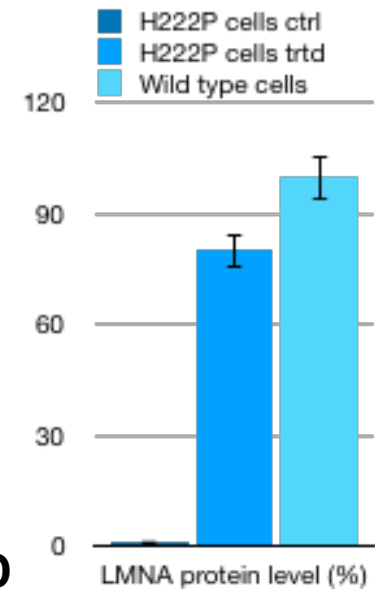
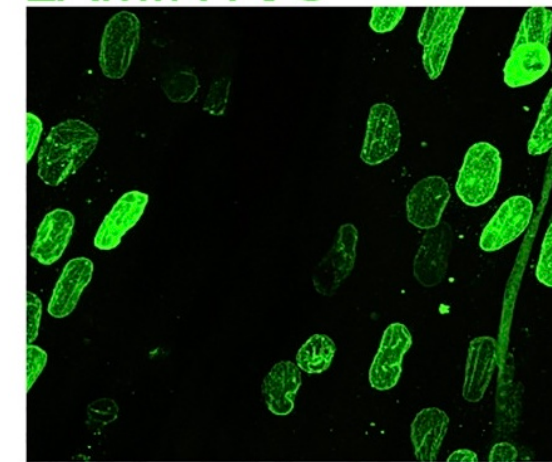


# In vitro experiment

# Analysis



## In H222P treated cells LAMIN A/C



# In vivo

4 months  
Sacrifice

1 day  
1<sup>st</sup> administration of  
gesicles

5 days  
2<sup>nd</sup> administration  
of lentivirus vector

110 days  
RotaRod test

## Timeline

## Materials

Lmna H222P/H222P mice

IN VIVO EXPERIMENTS

IN VIVO RESULTS

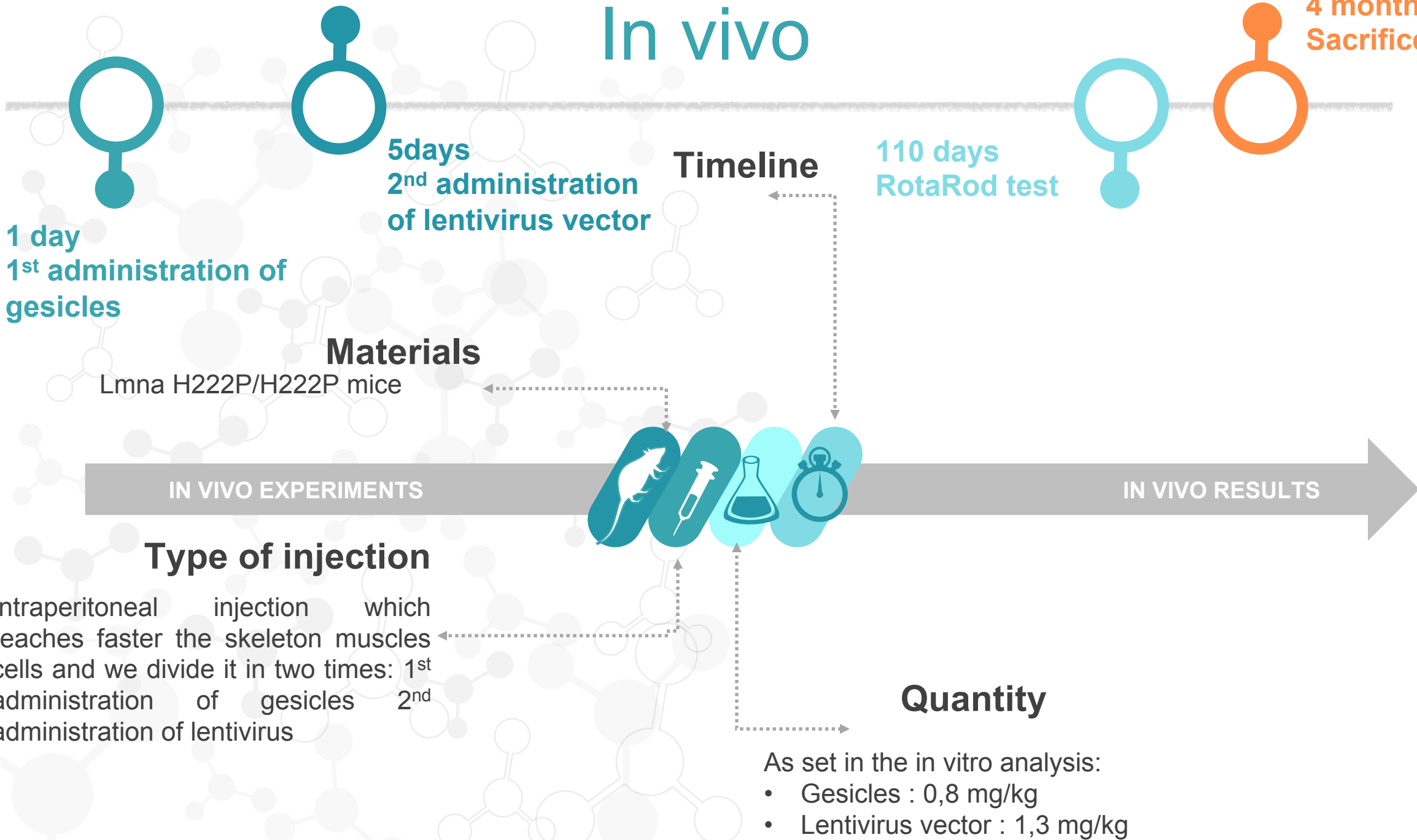
## Type of injection

Intraperitoneal injection which reaches faster the skeleton muscles cells and we divide it in two times: 1<sup>st</sup> administration of gesicles 2<sup>nd</sup> administration of lentivirus

## Quantity

As set in the in vitro analysis:

- Gesicles : 0,8 mg/kg
- Lentivirus vector : 1,3 mg/kg

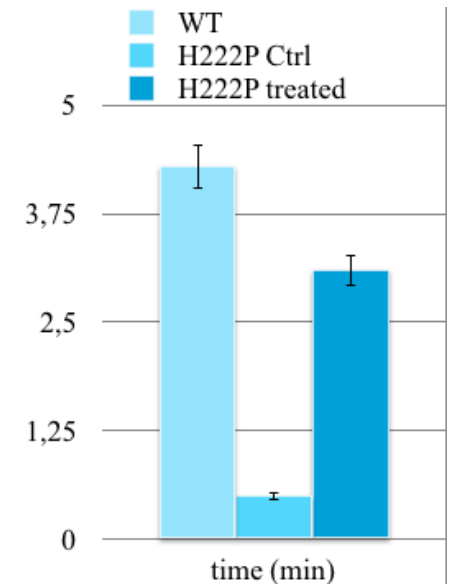


# In vivo experiment

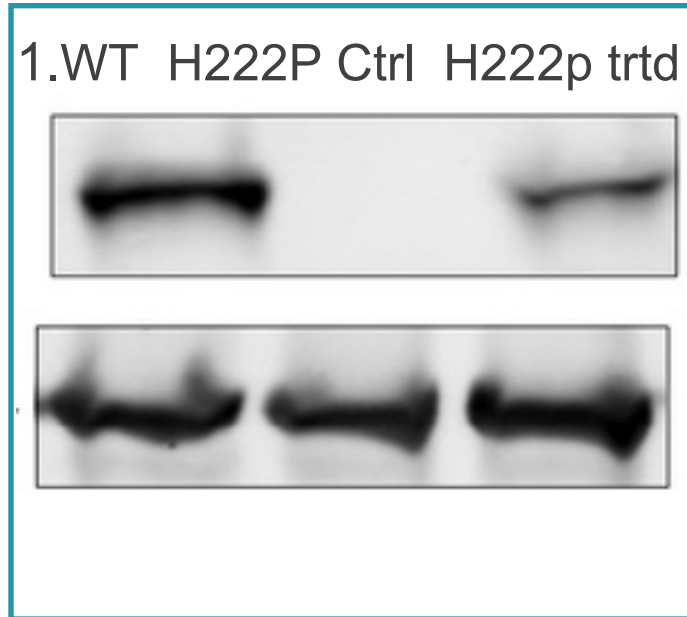
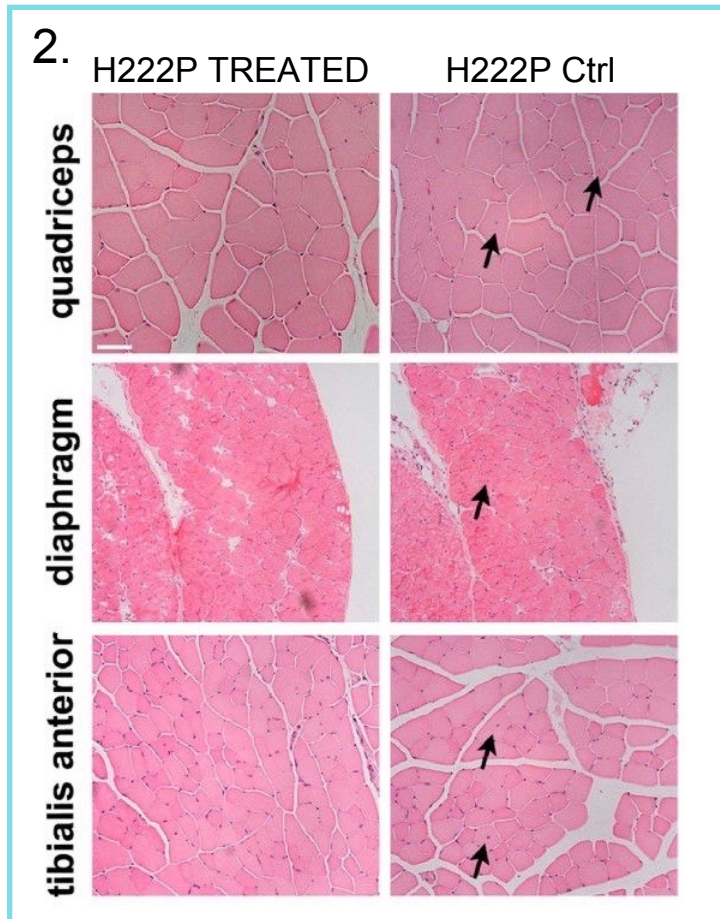
## results

3. RotaRod test is conducted after 110 days from birth and is used to evaluate the motor coordination of rodents.

3.



2. Histological images to show how the phenotype can be reverted



1. Western blot to analyze the production of LMNA after our treatment



# Materials & costs of production



1. Genscript CRISPR plasmid collection 500€
2. Cells cultures of skeletal muscle 500€
3. Immunofluorescence kit 520€ (ThermoFisher)
4. Lentiviral Packaging Mix: #VP100 400€ (AddGene)
5. Kit western Blot (reagents + instrumentation) 750€ (Thermo Fischer Scientific)
6. 30x (LMNA/H222P) mouse model 7200€/240€ each mouse (The Jackson Laboratory)
7. 15x (WT) mouse/1 free for each mouse model H222P (The Jackson Laboratory)
8. RotaRod test kit 320€ (PanLab)
9. Additional costs from basic lab maintenance and materials
10. Stabulation cost 1000€ each mouse for one year



Total cost : approximately  
**60.000€** without the salary  
cost of the researchers

# References

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