

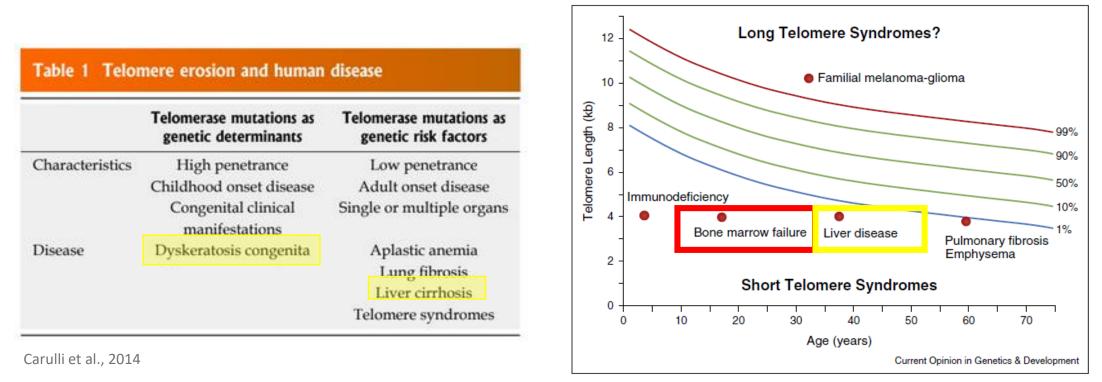
IMPROVE LIFE EXPECTATION OF PATIENTS AFFECTED BY TELOMEROPATHIES USING CD34+ CELL THERAPY

Muskaj, Spagnoli e Vastarelli

Class of Biology of stem cells, Prof. Isabella Saggio e Mattia La Torre

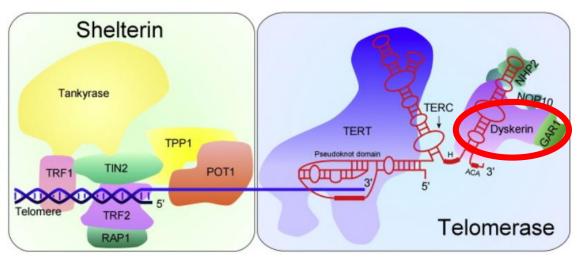
Telomere length and predominant clinical manifestations

Patients with <u>dyskeratosis congenita</u> (DC) suffer from stem cell failure in highly proliferative tissues. **DKC1** gene is the gene responsible for the X-linked Dyskeratosis Congenita.

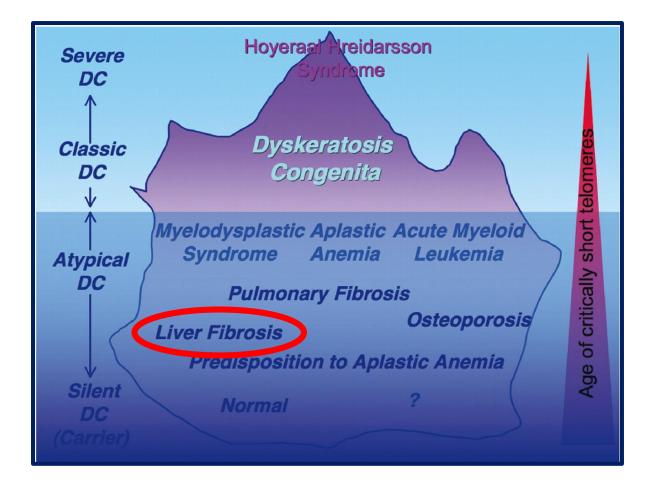


Armanios et al., 2015

Kirwan and Dokal, 2009

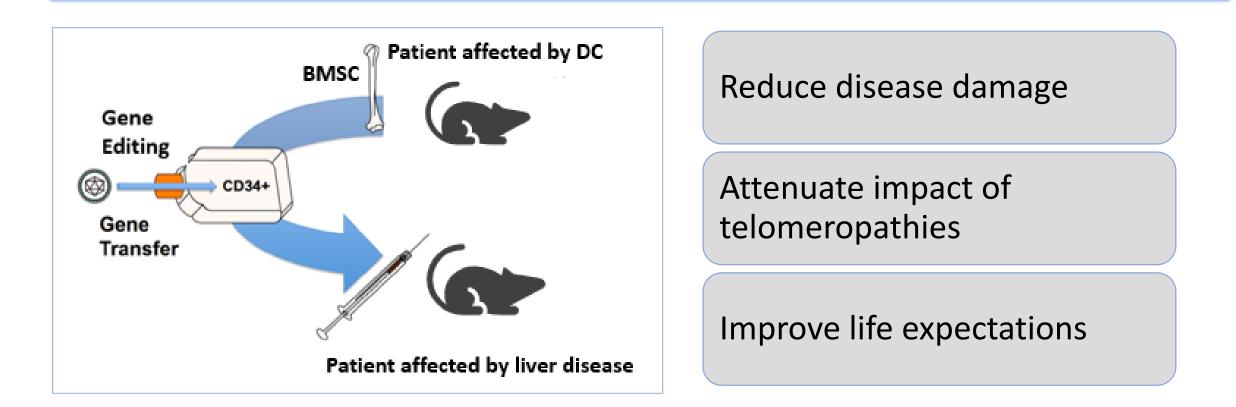


The **dysckerin complex** is a protein encoded by the gene DKC1. This cause a selective defect in the translation of a subgroup of internal ribosome entry site (IRES)–containing cellular mRNAs.



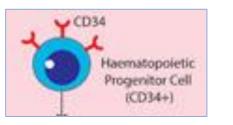
M. Bessler, Hong-Yan Du, Baiwei Gu, P. J.Mason, 2007

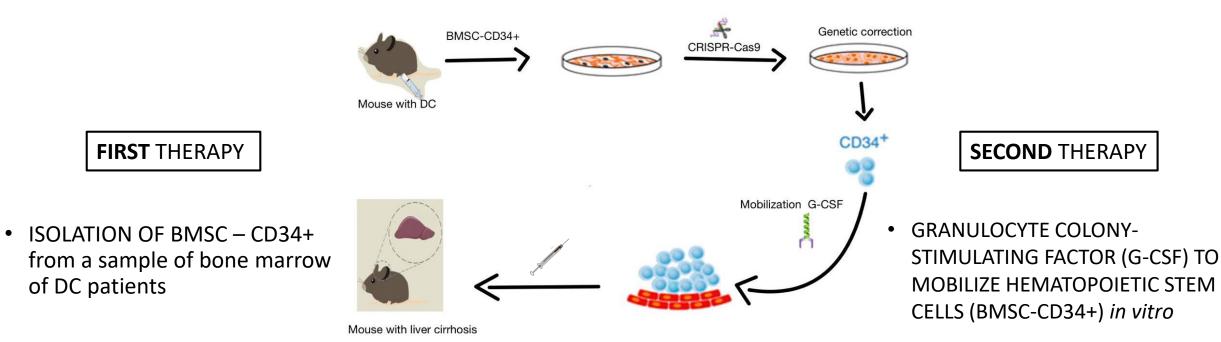
AIMs of the gene therapy



The short telomere phenotype in children and young adults represents more severe disease. Bone marrow failure is its most common first manifestation, and **stem cell transplantation** alleviates this condition pointing to a stem cell-autonomous defect in this compartment.

COMBINED THERAPY





• GENE EDITING with CRISPR *in vitro* to correct **DKC1** mutation

 TRANSPLANT *in vivo* OF CD34+ IN PATIENT AFFECTED BY LIVER DISEASE

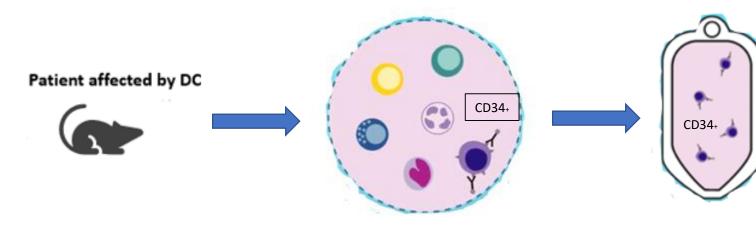
ISOLATION OF CD34+

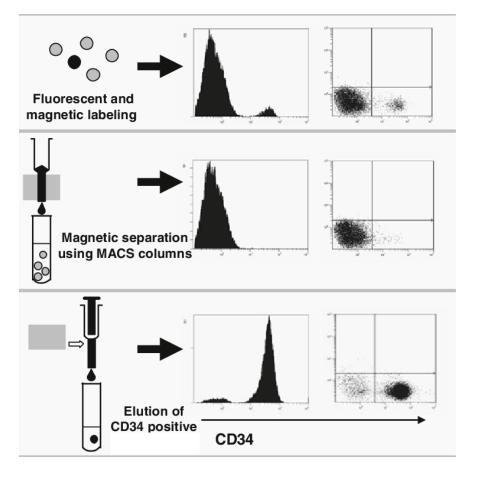


BONE MARROW SAMPLE FOR THE ISOLATION OF CD34+ FRACTION with CliniMACS® System

BM blood cell = $10.000/\mu$ L. \rightarrow CD34+ \rightarrow > 2 × $10^6/1$

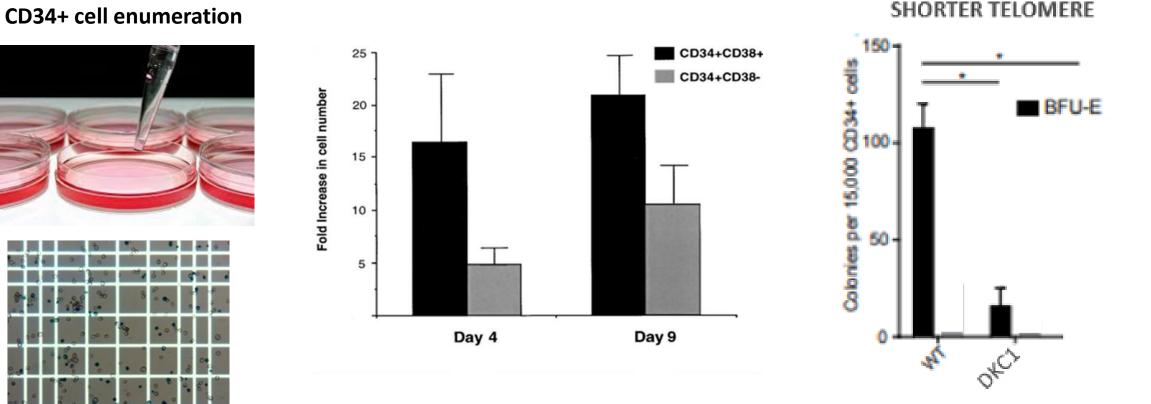
BMCs were sampled for cell counts and immunophenotyping by flow cytometry prior to processing.



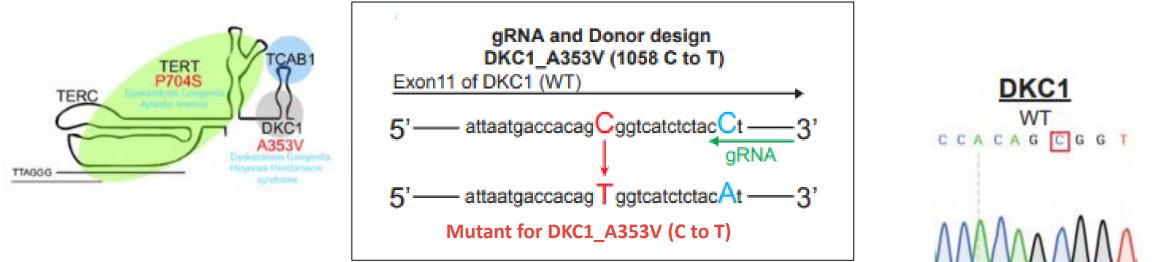


COLONIES OF CD34+

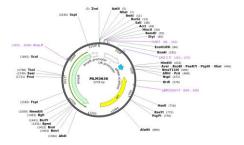
The percentage of CD34+ and viability were determined by flow cytometry analysis, used to calculate total CD34+ cells number.



CRISPR-Cas9 to correct DCK1 mutation



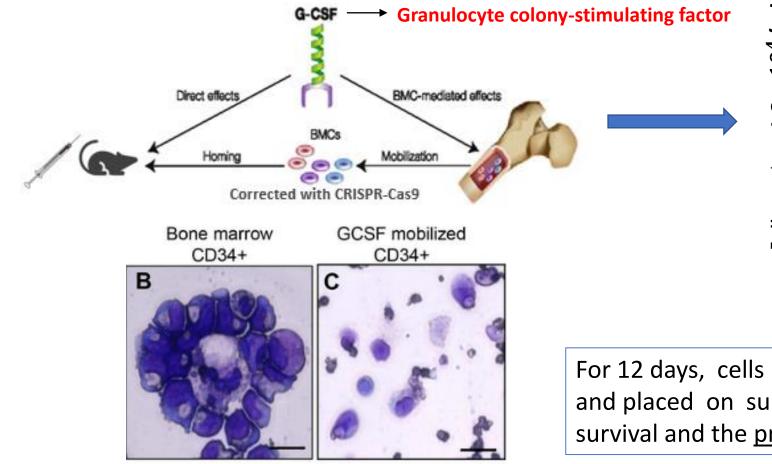
CRISPR gRNAs were inserted into the **MLM3636 plasmid** and cotransfected with a plasmid carrying Cas9.

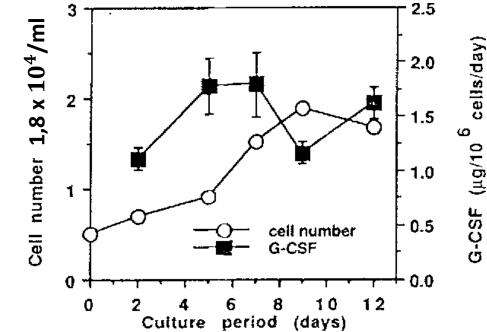


Sequencing traces confirming genome modification \rightarrow

А 1058C>T / АЗ5ЗV С С А С А G Т G G T М М 1100

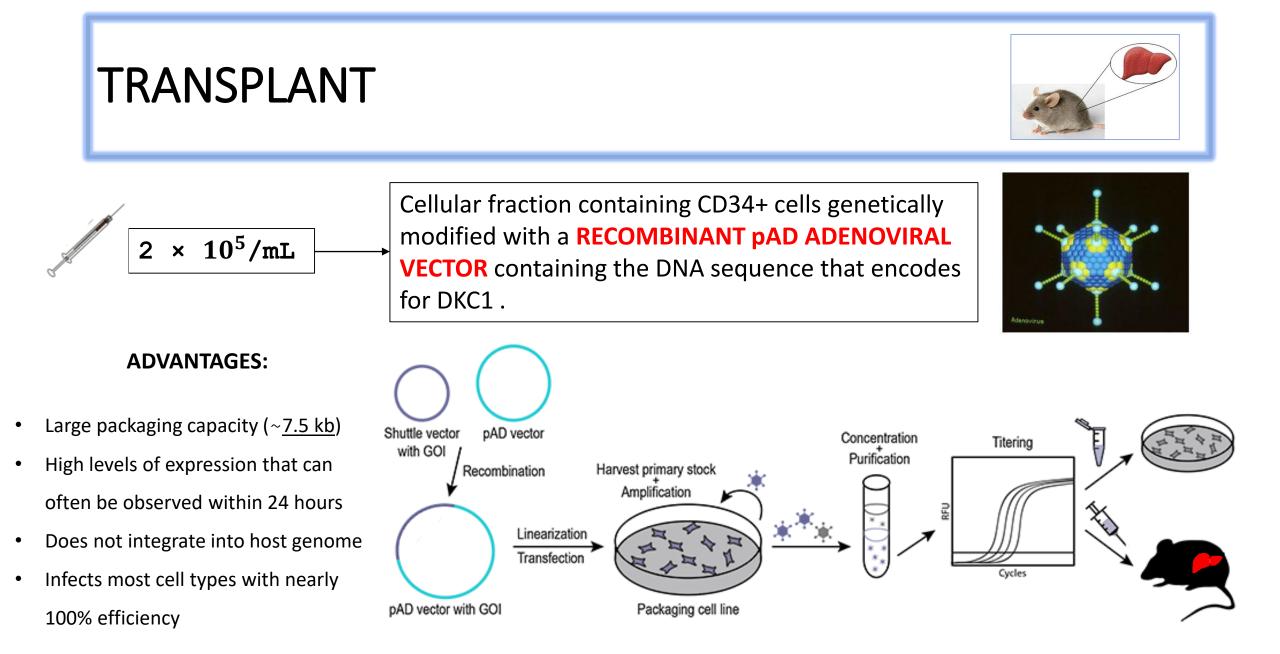
CD34+ cultured in G-CSF



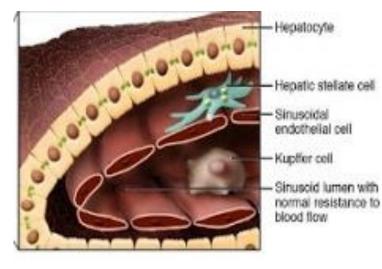


For 12 days, cells were cultured in **G-CSF**, washed and placed on superfrost slides, to stimulate the survival and the <u>proliferation</u>.

Methylthioninium chloride



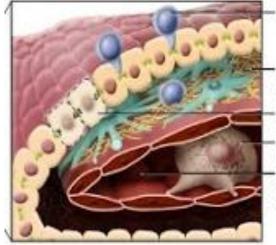
NORMAL LIVER



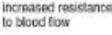
RESULTS

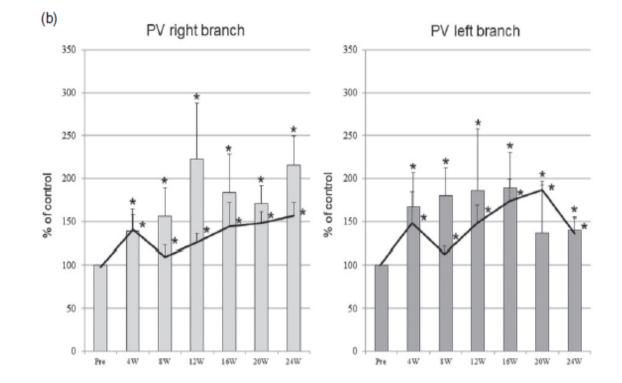
Improvement of liver function after CD34+ cell transplantation based on the analysis of **portal blood flow and velocity in both branches of the portal vein.**

LIVER WITH FIBROSIS

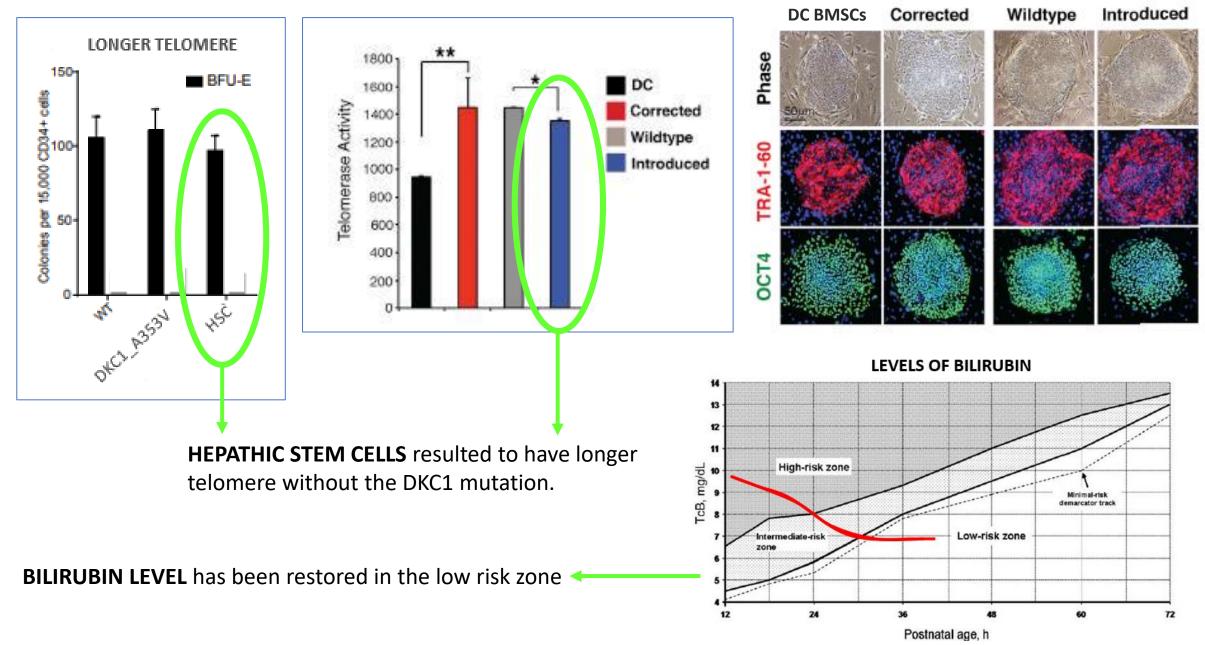


Infiltrating lymphocyte
 Extracellular matrix
 proteins
 Apoptotic hepatocyte
 Activated Kupfler cell
 Sinuscid lumen with





<u>Telomere length</u> quantified by Telomere Repeat Fragment Analysis (TRF).



Materials & budget

	20€ x 10 models = 200,00 €	1		
DKC1 mouse models	20€ X 10 models – 200,00 €	• M	ay trigger a s	sub
Liver cirrhosis mouse + WT mouse models	20€ x 30 models = 600,00 €		 Transient express Cloning can be ch Risk of hepathic t 	
CliniMACS [®] System TS 500 for Research Use	1.650,00 €			
CRISPR-Cas9 Mutation Detection Kit	160,00€			
G-CSF Recombinant Protein	500,00 €			30
pAD Adenoviral vector	1.200,00 €			
Additional costs (results analysis, markers,)	500,00 €		prrection of l	
Salary of researchers	3.500,00 €	te	lomeropathi	es
TOT.	8.310,00 €			
months Isolation and CRISPR gene editing 6 months	Proliferation and transplant 1 yea	ar II	<i>n vivo</i> resear	rch



- trigger a substantial immune response in vivo
- sient expression
- ing can be challenging due to large genome size
- of hepathic tumor



ection of liver disease mutation linked to neropathies

2 years

To be continued...

References



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THANKS FOR YOUR ATTENTION!